

One Team, One Purpose



Food Safety and Inspection Service

Protecting Public Health and Preventing Foodborne Illness



FSIS: Analytical Tools for WGS



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- Background
- What are we doing (or planning to do) with our data?
- Concluding remarks

Food Safety and Inspection Service: Why WGS?

- Improved resolution for foodborne illness investigations
 - Improved strain discrimination, illness cluster detection, and case classification

• Supports FSIS mission goals

- Effectively use science to understand foodborne illness and emerging microbiological trends
- Identification of environmental harborage or recurrences of pathogens in FSIS-regulated establishments/products to further support the inspection and verification process
- Alignment of pathogen surveillance with our domestic public health and regulatory partners
 - Collaborative efforts with US Food and Drug Administration Center for Food Safety and Applied Nutrition (FDA-CFSAN), the US Centers for Disease Control and Prevention (CDC), the US National Institutes of Health National Center for Biotechnology Information (NCBI), and also state/local health partners/laboratories

Food Safety and Inspection Service: WGS at FSIS: Current Status

- FSIS has built capacity for conducting WGS on all isolates obtained from FSIS sampling programs
 - Currently 12 sequencers in FSIS Field Service Laboratories, and expect all to be operational in early FY18
 - In FY17, FSIS sequenced 7,282 isolates
- In collaboration with our public health and regulatory partners, FSIS currently considers available WGS analyses in addition to PFGE, epidemiological and traceback information to further understand the relationship between clinical and food isolates
- FSIS works with National Antimicrobial Resistance Monitoring System (NARMS) partners (FDA, CDC) to understand the occurrence or introduction of antimicrobial resistance genes in pathogens of interest

Food Safety and Inspection Service: FSIS Sequencing Efforts and Data Sharing

FSIS Submissions to NCBI Bioprojects

- PRJNA242847
 - GenomeTrakr Project: USDA-FSIS (*Salmonella*)
- PRJNA215355
 - GenomeTrakr Project: FDA (Listeria monocytogenes)
- PRJNA287430
 - USDA-FSIS: Campylobacter
- PRJNA268206
 - GenomeTrakr Project: USDA-FSIS (STEC)
- PRJNA292666
 - FSIS NARMS Salmonella
- PRJNA292667
 - FSIS NARMS E. coli
- PRJNA292668
 - FSIS NARMS Campylobacter
- PRJNA292669
 - FSIS NARMS Enterococcus

Number of Isolates Sequenced by Fiscal Year



Food Safety and Inspection Service: WGS Data Sharing – Metadata

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ackage	Pathogen: environmental	/food/other; version 1.0		Search details	
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- Description about an isolate that has an experiment assigned
- Metadata, such as source type, organism, serotype etc.
- BioSample number is specific to a bacterial isolate for FSIS

Food Safety and Inspection Service: FSIS: WGS Analytical Tools Overview

- Quality Control Assessment
- Antibiotic Resistance gene detection
- Salmonella and STEC serotype determination
- STEC Virulence Gene characterization
- Phylogenetic Comparisons

Further Characterization of Isolates Using WGS

Campylobacter speciation Serotyping/serogrouping Salmonella Adulterant STEC Antimicrobial Resistance (Phenotype • prediction) Salmonella Campylobacter – E. coli Enterococcus Identify characterized genes of interest • Resistance to environmental factors (heat, acid, etc) Virulence factors • *stx/eae* sub-types (STEC) Alternative to PFGE for comparison of ۲ genotypes wgMLST analyses SNP analyses

A single workflow for many characterization

 approaches via informatics

Food Safety and Inspection Service: FSIS: WGS Data Analyses Work Flow Overview



Genotypic Screening for Antimicrobial Resistance Using WGS Data

- Ability to rapidly identify new genes of concern in sequenced isolates
- FSIS works with NARMS and other public health and regulatory partners to identify presence of genes or genetic markers of interest
 - When possible, FSIS currently compares genotype prediction to phenotype using the NARMS panel

2015

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Salmonella and

Campylobacter

antimicrobial

from cecal with

- Examples:
 - Extended spectrum betalactamases (*bla*_{CTX-M-65})
 - Colistin
 - Quinolone
 - Linezolid
 - Daptomycin

2017

and

All Salmonella

Campylobacter

from both cecal

AMR E. coli and Enterococcus

and HACCP

2016

and

All Salmonella

Campylobacter

from cecal

Food Safety and Inspection Service: WGS usage: Antibiotic Resistance Gene Detection



Food Safety and Inspection Service: WGS usage: Antibiotic Resistance Gene Detection

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Found 1 tetracycline genes in FSIS11704693.fa Processing: FSIS11704693.fa
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Found 0 vancomycin genes in FSIS11704693.fa



https://bitbucket.org/genomicepidemiology/resfinder

GENE	%COVERAGE	%IDENTITY	#FILE	CATEGORY	SEQUENCE
blaCTX-M-65	100	100	FSIS11704693	Beta-lactam resistance	FSIS11704693_S10_L001_R1_001_(paired)_trimmed_(paired)_contig_19
aadA1	95.37	99.89	FSIS11704693	Aminoglycoside resistance	FSIS11704693_S10_L001_R1_001_(paired)_trimmed_(paired)_contig_45
tet(A)	100	100	FSIS11704693	Tetracycline resistance	FSIS11704693_S10_L001_R1_001_(paired)_trimmed_(paired)_contig_45
sul1	100	100	FSIS11704693	Sulphonamide resistance	FSIS11704693_S10_L001_R1_001_(paired)_trimmed_(paired)_contig_45
floR	99.92	98.19	FSIS11704693	Phenicol resistance	FSIS11704693_S10_L001_R1_001_(paired)_trimmed_(paired)_contig_70

slide

Antimicrobial Resistance (Phenotype Prediction) Using WGS Analyses

- Analyzed ~1190 Salmonella isolates from FY2016 NARMS cecal sampling
 - Includes isolates from all animals tested
- Compared genotypic prediction for resistance phenotype to reported phenotype using NARMS panel
 - Overall concordance for 1130 isolates (95.0%)

Phenotype	Number of Isolates with Phenotype Using NARMS Panel	Percent Concordance of Genotype with Phenotype
Pan-susceptible	769	99.1%
Beta-lactam resistance	110	99.1%
Aminoglycoside resistance	167	98.8%
Phenicol resistance	42	97.6%
Tetracycline resistance	330	99.7%

Food Safety and Inspection Service: WGS usage: Salmonella Serotype Determination



Using WGS Analyses for Serotype Determination

- FSIS sequenced 4205 *Salmonella* isolates from various sampling programs from Jan 2015 through Jun 2017
- Compared serotype reported by routine methods (molecular serotyping or traditional serology) with serotype determined using a custom query to SeqSero database
 - For 4045/4205 (96.2%) of isolates, WGS that matched reported serology result
 - For 160/4205 (3.8%) of isolates, WGS did not match reported serology result
 - Includes isolates with incomplete genetic factor set (cannot call/identify serotype)

WGS usage: MLST and virulence typing in Shiga toxin-producing *Escherichia coli*



WGS usage: MLST and virulence typing in Shiga toxin-producing Escherichia coli

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FSIS_Number	Serotype	Sequence_type	stx_type	eae_allele
FSIS1608854	O26:H11	ST-21	stx1a	Beta1

Using WGS Analyses for MLST and virulence typing in Shiga toxin-producing *Escherichia coli*

Serogroup (no. sequenced)	Top 7 gene MLST sequence types	Top <i>stx</i> types	<i>eae</i> types	Top Serotype
O26 (50)	ST-21 (82%)	stx1a (80%) stx2a (16%)	Beta1 (100%)	O26:H11 (100%)
O45 (20)	ST-17 (95%)	stx1a (100%)	Epsilon (95%)	O111:H8 (100%)
0103 (147)	ST-17 (67.8%)	stx1a (98.0%)	Epsilon (84.9%)	O103:H2 (84.9%)
0111 (44)	ST-16 (93.0%)	stx1a (86.0%)	Theta (100%)	O111:H8 (100%)
0121 (15)	ST-655 (93.3%)	stx2a (100%)	Epsilon (100%)	O121:H19 (100%)
O145 (18)	ST-32 (100%)	stx1a stx2d (38.9%) stx2a (27.7%) stx2c (11.1%)	Gamma-1 (100%)	O145:H28 (100%)
0157 (149)	ST-11 (96.00%)	stx1a stx2a (28.2%) stx2c (22.8%) stx2a (20.1%)	Gamma-1 (100%)	O157:H7 (100%)

- *E. coli* serotypes, O157:H11 & O157:H29 identified to be lacking *eae/stx* through WGS
- *stx*-negative strains can be identified as serotype *E. coli* O157:H7 based on WGS

WGS Analyses for Phylogenetic Context: wgMLST and hqSNP Analyses

FSIS uses pipelines developed by public health partners Lyve-SET <u>https://github.com/lskatz/lyve-SET/blob/master/</u>

NCBI Pathogen Detection Isolates Browswer http://www.ncbi.nlm.nih.gov/pathogens

FDA SNP Pipeline https://github.com/CFSAN-Biostatistics/snp-pipeline

wgMLST BioNumerics 7.6 CDC-PulseNet



Food Safety and Inspection Service WGS usage: SNP tree from NCBI Pathogen Pipeline

https://www.ncbi.nlm.nih.gov/pathogens/isolates/

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Product	Serotype	Total #	% in SNP cluster (#)	% w/in 20 SNPs of clinical	% w/in 10 SNPs of clinical
Total Serotypes	Enteritidis	447	99.8% (446)	98.9% (442)	90.6% (405)
	Kentucky	607	99.8% (606)	0% (0)	0% (0)
	Heidelberg	74	98.6% (73)	98.6% (73)	86.4% (64)
	Schwarzengrund	122	100% (122)	47.5% (58)	7.3% (9)
	Infantis	218	94.5% (206)	92.2% (201)	50.9% (111)
	Typhimurium	155	96.8% (150)	30.3% (47)	10.3% (16)

Food Safety and Inspection Service WGS usage: SNP tree from NCBI Pathogen Pipeline



Food Safety and Inspection Service WGS usage: hqSNP analyses and wgMLST with *Listeria monocytogenes*



Food Safety and Inspection Service WGS usage: hqSNP analyses and wgMLST with *Listeria monocytogenes*

Lab Number	Primary_PN_Pattern	Sec_PN_Pattern	Lyve-SET SNP Clade	CFSAN Snp Pipeline Clade	wgMLST Clade
FSIS1400040	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1400020	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1400249	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1400248	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1400246	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1500681	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1500682	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1500683	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1500684	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1500685	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1500686	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1500688	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1500689	GX6A16.0018	GX6A12.0022	1	l .	1 1
FSIS1500690	GX6A16.0018	GX6A12.0022	1	l .	1 1
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FSIS1502845	GX6A16.0018	GX6A12.0022	ź	2	2 <mark>22</mark>
FSIS1502847	GX6A16.0018	GX6A12.0022	ź	2	2 <mark>22</mark>
FSIS1502848	GX6A16.0018	GX6A12.0022	2	2	2 <mark>22</mark>
FSIS1502849	GX6A16.0018	GX6A12.0022	2	2	2 2
FSIS1502850	GX6A16.0018	GX6A12.0022	2	2	2 2
FSIS1502851	GX6A16.0018	GX6A12.0022		2	2 2

<mark>0-5</mark>	
0-10	
0-15	
0-20	

WGS usage: Usage of genotypic data to predict phenotypic characteristics

- FSIS is beginning to explore using genotypic data to predict phenotypic characteristics
 - Previous examples of antimicrobial resistance markers
 - Resistance to other environmental factors (i.e. heat, acid, certain chemicals, etc)
- Example: Locus of Heat Resistance (LHR) 14-kb genomic island
 - Present in a diverse group of Enterobacteriaceae, including Cronobacter sakazakii, Klebsiella pneumoniae, Enterobacter cloacae, E. coli, and Salmonella
 - A BLAST database including genes contained in the LHR was built based on the published sequence of *E. coli* AW1.7
 - Salmonella isolates from FSIS-regulated products were sequenced, assembled and queried against the LHR BLAST database
 - 11 FSIS *Salmonella* isolates contained LHR genes of interest
 - Additional phenotypic testing required to determine if isolates exhibit resistance to heat
- Heat resistance can be linked to different biological pathways/genes
 - Advantage of WGS: As new pathways/genes are identified, data can be queried again



Concluding Remarks

- FSIS' new strategic plan is focused on the use of new technology to prevent foodborne illnesses and protect public health
 - Includes two objectives specific to incorporating WGS into FSIS surveillance and regulatory efforts
- FSIS has built sufficient capacity for conducting WGS on all FSIS pathogen isolates
 - Sequenced 7282 isolates in FY17 with the future target of ~10,000 isolates/year
- FSIS is exploring how we can use WGS data beyond outbreak investigations, including understanding the link between genotypes and phenotypes of interest
- FSIS continues to engage with national and international partners
- FSIS continues to use WGS analyses in conjunction with other metadata, including epidemiological and traceback information, to further understand the relationship between clinical, food and environmental isolates

Food Safety and Inspection Service: Acknowledgements



National Center for Biotechnology NCBI Information

- USDA FSIS Offices
- USDA ARS
- CDC PulseNet and NARMS
- FDA CFSAN
- FDA CVM
- NCBI
- State Laboratories











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