I. PURPOSE

This directive provides instructions to Public Health Veterinarians (PHVs) at slaughter establishments selected for sampling for the National Antimicrobial Resistance Monitoring System (NARMS) surveillance sampling program. For the purpose of this directive, the term “PHV” refers to both PHVs and Supervisory Public Health Veterinarians (SPHVs). This directive issues instructions from FSIS Notice 13-13 and includes revised instructions on ordering sampling supplies. PHVs are to order NARMS sampling supplies at least 72 hours before the scheduled sample collection date. These instructions are consistent with the instructions in other sampling notices and directives.

KEY POINTS:

- PHVs are to collect samples of cecal contents from the large intestines of select classes of swine, cattle, and poultry in FSIS-regulated establishments for the NARMS sampling program
- NARMS sampling tasks are assigned to PHVs at a frequency based on the animal classes slaughtered and the establishment’s annual slaughter volumes, using information entered in the Public Health Information System (PHIS)
- NARMS sampling tasks are assigned solely to PHVs; PHVs complete these tasks through PHIS
- Sample results from the NARMS sampling program are non-regulatory; therefore, establishments do not need to hold products
- Results of individual samples collected will not be posted in Laboratory Information Management System (LIMS)-Direct or PHIS

II. BACKGROUND

A. NARMS is a national public health surveillance system that tracks antimicrobial susceptibility among enteric bacteria from humans, retail meats, and food animals. This program was established in 1996 as a partnership between the U.S. Food and Drug Administration (FDA), the Centers for Disease Control and Prevention (CDC), and FSIS. In addition to monitoring antimicrobial susceptibility, the NARMS partners collaborate on epidemiologic and microbiologic studies and conduct research to better understand the emergence, persistence, and spread of antimicrobial resistance among foodborne bacteria.

B. Purpose of the sampling. The FSIS NARMS sampling program provides data on the presence and
antimicrobial resistance profile of selected enteric microorganisms in food animal species. As part of this sampling program, the Office of Public Health Science (OPHS) laboratory in Athens, Georgia will process samples of cecal contents collected by PHVs and test for the presence of *Salmonella*, generic *Escherichia coli* (*E. coli*), *Campylobacter*, and *Enterococcus* spp. FSIS will perform the primary isolation and identification of these microorganisms, and will complete further characterization of the isolates to include serotyping, pulsed field gel electrophoresis (PFGE), and antimicrobial susceptibility testing. The resulting data will be used in NARMS to monitor trends in antimicrobial resistance and susceptibility among enteric bacteria in food animals.

C. **Sampling program description.** PHVs will collect samples of cecal contents from the large intestines of swine (Market Swine, Sow), cattle (Dairy Cow, Beef Cow, Steers, and Heifers), Young Chickens, and Young Turkeys in FSIS-regulated livestock and poultry slaughter establishments. Samples collected will be shipped to the FSIS Eastern Laboratory (EL) for testing, as described in Section X.

### III. ESTABLISHMENT ELIGIBILITY FOR THE FSIS NARMS SAMPLING PROGRAM

A. Livestock and poultry slaughter establishments are eligible for the NARMS sampling program based on data in PHIS: by establishment size; the animal classes slaughtered; and annual slaughter volumes. Sampling tasks for the NARMS sampling program will be assigned at the following frequencies, using the previous 12 months of slaughter data:

<table>
<thead>
<tr>
<th>Slaughter Volume</th>
<th>Maximum number of sampling tasks per month per sampling project</th>
</tr>
</thead>
<tbody>
<tr>
<td>Top 25% of Slaughter Establishments</td>
<td>4</td>
</tr>
<tr>
<td>Second 25% of Slaughter Establishments</td>
<td>2</td>
</tr>
<tr>
<td>Lowest 50% of Slaughter Establishments</td>
<td>1</td>
</tr>
</tbody>
</table>

B. PHVs at these establishments will receive sampling tasks through PHIS and are to order supplies for each sampling event from the EL (see Section VI). PHVs are to respond to each sampling task using the instructions provided in this directive.

### IV. REFERENCES AND REVIEW OF TRAINING MATERIALS

A. PHVs assigned to establishments eligible for the FSIS NARMS sampling program are to be familiar with the following FSIS issuances and the information provided in this directive:

1. **PHIS Directive 13000.2**, *Performing Sampling Tasks in Official Establishments Using the Public Health Information System*

2. **FSIS Directive 7355.1, Revision 2**, *Use of Sample Seals for Laboratory Samples and Other Applications*

3. **NARMS Training video**

B. PHVs who are unable to access the training video through the FSIS intranet site are to contact their supervisor and request a copy of the video on DVD.

### V. SAMPLING TASK ASSIGNMENT

A. Notification of sampling tasks will be in the form of an alert in PHIS. The sampling tasks will be
directed tasks on the establishment task list with one or more of the following sampling project codes:

<table>
<thead>
<tr>
<th>Project Code</th>
<th>Slaughter Class To Sample</th>
</tr>
</thead>
<tbody>
<tr>
<td>NARMS_YC</td>
<td>Young Chicken</td>
</tr>
<tr>
<td>NARMS_YT</td>
<td>Young Turkey</td>
</tr>
<tr>
<td>NARMS_DC</td>
<td>Dairy Cow</td>
</tr>
<tr>
<td>NARMS_BC</td>
<td>Beef Cow</td>
</tr>
<tr>
<td>NARMS_ST</td>
<td>Steer</td>
</tr>
<tr>
<td>NARMS_HF</td>
<td>Heifer</td>
</tr>
<tr>
<td>NARMS_MS</td>
<td>Market Swine</td>
</tr>
<tr>
<td>NARMS_SW</td>
<td>Sow</td>
</tr>
</tbody>
</table>

B. The sampling task can be located on the task list in PHIS by using the “sampling” filter. The PHV is to refer to FSIS Directive 13000.2 for instructions on how to add the task to the task calendar, enter the sample information, submit the sample information to the lab, and print a finalized sample collection form in PHIS.

C. The sample collection window for each NARMS sampling project is 37 days. The PHV is to schedule sample collection as soon as possible to reserve laboratory capacity. Samples cannot be scheduled once the laboratory has met its daily testing capacity.

VI. ORDERING SAMPLING SUPPLIES

A. The PHV is to request NARMS sampling supplies necessary for the slaughter class scheduled for sampling at least 72 hours before the scheduled sample collection date. The EL will not automatically send sampling supplies. The PHV is to use only the sampling supplies provided by the EL that are specific to slaughter class scheduled for sample collection.

B. The PHV is to refer to the Sampling Supply List in Section VIII.A. for a list of sampling supplies provided by the EL. These supplies do not include isopropyl alcohol, isopropyl alcohol wipes or pads, or a caddy, which must be procured locally. The PHV is to contact the Frontline Supervisor (FLS) for guidance on obtaining these supplies.

C. The PHV is to order sampling supplies through the PHIS task calendar. To order sample supplies, PHVs are to right-click on the scheduled NARMS sampling task and select Order Supplies from the dropdown menu. A pop-up window will appear that displays the project code and the name of the FSIS Laboratory that will fill the sampling supply request. When necessary, PHV are to enter requests for specific supplies (e.g., extra gloves) in the Comments field and click Submit Request. A confirmation message will appear.

D. As an alternative in situations where the PHV is unable to order supplies through the task calendar (e.g., because of PHIS connectivity issues), the PHV may send a request for sampling supplies using Outlook to the FSIS - Sampling Supplies – Eastern Lab mailbox.

E. The PHV is to use the Subject heading “NARMS Sampling Supplies” in the email and include the establishment name and number; the project code; the PHV’s contact name and telephone number; and a list of the supplies needed.
VII. SAMPLE SELECTION

A. The PHV is to refer to the NARMS sampling task for information on the species and slaughter class to be sampled.

B. The PHV is to randomly select from the lots of animals presented for slaughter on the scheduled sampling day that have passed ante-mortem inspection. The PHV is to select a carcass post mortem for each scheduled sampling event. The PHV is to note the lot information and animal identification information for the animals selected and record this information in the appropriate data fields in the Sample Collection screen in PHIS for the sampling task.

C. The PHV is to work with establishment management to identify:

1. The point in the slaughter process where the viscera / large intestines will be retrieved for sampling;
2. The location in the establishment where the sample selection will be performed;
3. The establishment employee designated to retrieve the selected viscera and transport it to the predetermined sample collection location for sample collection in livestock slaughter establishments; and
4. A cleanable work surface (i.e., table) in the sample collection location for staging the sampling supplies.

D. The number of cecal samples to collect per sampling task is as follows:

<table>
<thead>
<tr>
<th>Project Code</th>
<th>Slaughter class</th>
<th>Number of cecal samples</th>
</tr>
</thead>
<tbody>
<tr>
<td>NARMS_YC</td>
<td>Young Chicken</td>
<td>5 (composited)</td>
</tr>
<tr>
<td>NARMS_YT</td>
<td>Young Turkey</td>
<td>5 (composited)</td>
</tr>
<tr>
<td>NARMS_DC</td>
<td>Dairy Cow</td>
<td>1</td>
</tr>
<tr>
<td>NARMS_BC</td>
<td>Beef Cow</td>
<td>1</td>
</tr>
<tr>
<td>NARMS_ST</td>
<td>Steer</td>
<td>1</td>
</tr>
<tr>
<td>NARMS_HF</td>
<td>Heifer</td>
<td>1</td>
</tr>
<tr>
<td>NARMS_MS</td>
<td>Market Swine</td>
<td>1</td>
</tr>
<tr>
<td>NARMS_SW</td>
<td>Sow</td>
<td>1</td>
</tr>
</tbody>
</table>

E. The PHV is to perform sample collection in a location and in a manner that ensures their personal safety, maintains sanitary conditions, prevents the contamination of edible product, and does not interfere with inspection activities. Examples of sampling locations may include, but are not limited to, the veterinary disposition area on the kill floor or the intestine harvesting area (in an establishment that harvests intestines for edible purposes).

VIII. SAMPLE COLLECTION

A. PREPARATION FOR SAMPLE COLLECTION (all species)

NOTE: The PHV is responsible for collecting the samples and is not to delegate this task to a Consumer Safety Inspector (CSI) or any establishment employee.
Sampling Supplies provided by the FSIS Laboratory:

- Shipping container
- Sterile 120 ml plastic screw-top sample containers (2)
- Gallon-size zipper lock bag (1)
- Quart-size zipper lock bag (1)
- Permanent marker (1)
- Scissors (1 pair for poultry sample collection, 2 pairs for cattle and swine sample collection) (to be returned to lab with the sample)
- Non-sterile examination gloves (8)
- Tongue Depressors (1 per sample plus 2 extra)( Livestock samples only)
- 2’ × 3’ plastic pad (1)
- Sterile gauze pads (2)
- 60 cc catheter tipped syringe ( with protective cap) (Livestock samples only)
- 6” × 12” plastic sleeve for sampling form
- FSIS Form 7355-2A/2B (sample seals)
- Absorbent pad for shipping container
- Cardboard separator(s)
- Gel coolant pack (1)
- FedEx airbill (pre-printed)

Supplies Locally Procured - submit reimbursement Form 1164 to District Office.

- Bottle of 70% Isopropyl Alcohol
- Isopropyl alcohol wipes or pads
- Caddy or tote - if needed
Upon receipt of the sampling supplies:

1. Verify receipt of all supplies needed to perform the sample collection.
2. Remove gel coolant packs from the shipping container and place them in the freezer at least 24 hours prior to sample collection. Pre-chill the shipping container.

On the day of sample collection:

1. Review the sampling steps for the applicable species.
2. Take supplies to the pre-determined sample collection location using a caddy or tote.
3. If needed, wipe down the table that will be used as the work surface.
4. Lay out the plastic pad on the work surface to provide a clean surface and set out the sampling equipment.

NOTE: The type of scissors provided by the FSIS Laboratory may vary and may not be the same as those shown in the sampling instruction photos.

5. Label the sample container and quart-size zipper lock bag with the form number to match the sample with the sample submission form.

6. Loosen the screw cap on the sample container but do not remove it. Set it on the plastic pad.
7. Lay down a sterile gauze pad on the plastic pad. Carefully clean the tips and blades of scissors by using one or more alcohol wipes and position the blades of the scissors on the sterile gauze pad.

For cattle and swine sample collection, repeat this cleaning step on the second pair of scissors provided.

8. Wash hands with soap and water. Don two pairs of gloves in an aseptic manner.

B. POUlTRY SAMPLE COLLECTION (Young Chicken and Young Turkey)

1. Randomly select a set of viscera with intact ceca at the pre-determined sample selection location and take it to the sample collection location. If the selected viscera set is visibly contaminated with feed, ingesta, feces, bile, etc., select the next uncontaminated viscera.
2. Rinse gloves with 70% isopropyl alcohol.

**NOTE:** If one or both pairs of gloves become contaminated during initial collection of the viscera, remove the outer pair of gloves before rinsing the gloves.

3. With one hand, hold both ceca about 1" below their attachment to the ileum. This hand will be stationary and will hold the ceca throughout the procedure. With the other hand, carefully strip away the mesentery from the ceca without separating them from their attachment to the ileum. Milk the contents of the ceca down towards the apex using the same hand.

4. Gently wipe around the sphincter of the cecum using an alcohol wipe or pad.
5. Detach both ceca from their attachments to the small intestine by cutting through the sphincter area of each.

6. Place both ceca into the plastic specimen container without draining them.

**Note for Turkey Sampling:**

Both ceca are to be submitted without draining them. However, if the ceca are too large to fit into the sample container, drain the cecal contents into the sample container as follows:

a. Milk the contents to the blind pouches.

b. Rinse the end of the ceca with alcohol and wipe off excess alcohol with sterile gauze.

c. Position the ceca over the open collection bottle and carefully cut a small opening in the end of the ceca and let contents pour into the collection bottle. Repeat this process for the second ceca.

7. Repeat steps 1 – 6 until samples have been collected from 5 separate carcasses.

**NOTE:** The samples collected from each carcass are to be placed in the same sample collection container and submitted as a composited sample.
8. Replace the sample container lid and close tightly.

9. When sample collection is completed, wipe the scissors clean with alcohol wipes. Wrap the scissors in several layers of paper towels and place them into the shipping container with the sample.

   NOTE: The scissors must be returned to the FSIS Laboratory in the shipping container with the sample.

10. Discard the used disposable sampling supplies (wipes, plastic pad, and exam gloves) into an appropriate trash receptacle.

C. CATTLE SAMPLE COLLECTION (Beef Cow, Dairy Cow, Steer, Heifer)

1. Randomly select an intact intestine without ingesta or bile contamination at the predetermined sample selection location. If the randomly selected intestine is visibly contaminated with feed, ingesta, feces, bile, etc., select the next uncontaminated one.

2. Direct the randomly selected intestine to the sample collection location (See Section VII.C.3)

3. Grasp the apex of the cecum with one hand and detach it by gently breaking down connective tissue with your other hand. Scissors may be needed to cut through the connective tissue. Continue to grasp the cecum with the same hand throughout the sample collection.

4. Elevate the apex to allow the contents to flow into the main body of the cecum.
5. Rinse gloves with 70% isopropyl alcohol. Note: If one or both pairs of gloves become contaminated during initial collection of the viscera, remove the outer pair of gloves before rinsing the gloves.

6. Pour isopropyl alcohol over the apex of the cecum at the selected incision site. Let sit for 10 seconds.

7. Wipe area once with a sterile gauze pad.

8. Using scissors, make a 3-inch longitudinal incision into the cecum so that the cecal contents are easily visible.

   Use the second pair of scissors provided with the sample if the first pair becomes contaminated or is dropped during the course of sample collection.
9. Collect at least 30 ml. of cecal contents using one of the following methods, depending upon cecal content consistency:

**If the contents are a pasty consistency:**
Insert a tongue depressor into the cecum, withdraw cecal contents, and place the cecal contents into the sample container. Repeat 3-5 times until at least 30 ml of cecal contents is collected.

**If the contents are a watery consistency:**
Position the cecum over the edge of the sampling area. Using a non-bargaining unit assistant, position the open sample container below the cecum and direct the incision area downward to allow the cecal contents to flow out of the incision site and into the container. Cecal contents should flow freely into the specimen container while allowing you to control flow and prevent spillage.

**Do not allow the cecum to come in direct contact with the inner surface of the sample container.**

**An alternate sample collection option for contents of a watery consistency:**
Insert the tip of a 60cc catheter tipped syringe through the incision site and into the cecum. Draw back on the plunger of the syringe and withdraw at least 30 ml of cecal contents into the syringe. Slowly depress the contents of the syringe into the sample container.

**Do not allow the tip of the syringe to come in direct contact with the inner surface of the sample container.**

**NOTE:** This is a good option for collecting the sample when an assistant is not available.
10. Replace the sample container lid and close tightly. The container should be ¼ to ½ full.

11. When sample collection is completed, wipe the scissors clean with alcohol wipes. Wrap both scissors in several layers of paper towels and place them into the shipping container with the sample.

**NOTE:** Both scissors must be returned to the FSIS Laboratory in the shipping container with the sample.

12. Discard the disposable used sampling supplies (wipes, plastic pad, tongue depressor [if used], 60 cc syringe [if used], and exam gloves) into an appropriate trash receptacle.

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### D. SWINE SAMPLE COLLECTION (Market Swine and Sows)

1. Randomly select an intact intestinal tract at the pre-determined sample selection location. If the selected intestinal tract is visibly contaminated with feed, ingesta, feces, or bile, select the next one that is not contaminated.

2. Direct the randomly selected intestine to the sample collection location. (See Section VII.C.3).

3. Rinse gloves with 70% isopropyl alcohol.

**NOTE:** If one or both pairs of gloves become contaminated during initial collection of the viscera, remove the outer pair of gloves before rinsing the gloves.
4. Grasp the apex of the cecum with one hand and detach it by gently breaking down connective tissue with your other hand. Scissors may be needed to cut through the connective tissue. Continue to grasp the cecum with the same hand throughout the sample collection.

5. Elevate the apex to allow the contents to flow into the main body of the cecum.

6. Pour isopropyl alcohol over the apex of the cecum at the proposed incision site. Let sit for 10 seconds and wipe once with a clean gauze pad.

7. Using scissors, make a one-inch incision into the apex of the cecum along a cecal band.

**NOTE:** Use the second pair of scissors provided with the sample if the first pair becomes contaminated or is dropped during the course of sample collection.
8. Position the main body of the cecum over the edge of the table. Position the open sample container below the cecum.

(Note: the sample container may be held by a non-bargaining unit assistant or positioned on a lower stationary surface).

9. Carefully direct the incision over the sample container to catch the cecal contents and prevent spillage of contents onto the floor. Slowly tilt the cecum downward to allow cecal contents to flow freely into the specimen container.

Collect at least 30 ml of cecal contents.

If you are unable to obtain enough cecal content through pouring, use your free hand to milk the cecal contents out of the cecum, through the incision area, and into the sample container.

Do not allow the cecum to come in direct contact with the inner surface of the sample container.

Another option for sample collection if a non-bargaining unit assistant is not available:

Insert the tip of a 60cc catheter tipped syringe through the incision site and into the cecum. Draw back on the plunger of the syringe and withdraw at least 30 ml of cecal contents into the syringe. Slowly depress the contents of the syringe into the sample container.

Do not allow the tip of the syringe or catheter to come in direct contact with the inner surface of the sample container.

10. Replace the sample container lid and close tightly. The sample container should be ¼ to ½ full.
11. When sample collection is completed, wipe the scissors clean with alcohol wipes. Wrap both scissors in several layers of paper towels and place them into the shipping container with the sample.

**NOTE:** Both scissors must be returned to the FSIS Laboratory in the shipping container with the sample.

12. Discard the disposable used sampling supplies (wipes, plastic pad, tongue depressor [if used], 60 cc syringe [if used], and exam gloves) into an appropriate trash receptacle.

**IX. COMPLETING THE SAMPLING TASK IN PHIS**

A. The PHV is to follow the instructions provided in *FSIS Directive 13,000.2* and in the *PHIS User Guide* for accepting, scheduling, and completing the sampling task using PHIS. To assist in the sampling task, the PHV may choose to print a copy of the sample form from PHIS for use during sample collection and to document lot information to record in PHIS.

B. The PHV is to enter lot information, the owner/producer name and address, and animal identification information into the appropriate Sample Collection data fields in PHIS for each sample collected. The PHV is to ensure that all requested information is entered into PHIS. When sample collection data entry is completed, the PHV is to click the “Submit to Lab” button, print a finalized form, and sign and date the form. PHIS will display a message stating that the sample collection information has been successfully submitted. The PHV is to place the signed sample form in the sample box with the corresponding sample.

**X. SHIPPING THE SAMPLE**

A. The PHV is to use only the shipping materials provided by the FSIS laboratory specific to the NARMS sampling program and refer to *FSIS Directive 7335.1, Use of Sample Seals for Program Samples and Other Applications*, for complete instructions on the proper use of sample seals.

B. For samples collected from Monday to Wednesday morning, the PHV is to ship the sample no later than Wednesday afternoon to ensure its arrival at the FSIS Eastern Laboratory before Friday. For samples collected between Wednesday afternoon and Friday afternoon, IPP are to store and maintain the sample under refrigeration and under FSIS control and ship on Monday morning. Do not freeze the samples.

C. Upon completion of the sample collection, the PHV is to perform the steps below.

1. Apply one small barcode label from FSIS Form 7355-2A/2B, Laboratory Sample Container Seal Set, to the sample collection container.

2. Place the sample collection container in a quart-size zipper lock bag. Expel excess air from the bag and close the bag using the zipper lock closure. Place the bagged specimen into the gallon-size zipper lock bag, expel the excess air from the bag, and close the bag using the zipper lock closure.
3. Apply the medium sized bar-coded FSIS Laboratory Sample Identification Label (FSIS Form 7355-2B) to the gallon-size zipper lock bag.

4. Affix one small bar-coded sample label from FSIS Form 7355-2A/2B sample seal set to the completed and signed printed sample form.

5. Affix the bar-coded label in the space provided at the top center of the sampling form.

6. Place the completed sample form in the plastic sleeve provided.

7. Retrieve the frozen gel coolant packs from the freezer and retrieve the pre-chilled shipping container.

8. Place the cardboard separator and absorbent pad on the bottom of the shipping container.

9. Place the frozen gel coolant pack on top of the absorbent pad and then place the corrugated cardboard pad on top of the frozen gel coolant pack. Place the bagged sample on top of the corrugated cardboard pad.

**NOTE:** When needed, place a second frozen gel coolant pack in the shipping container to ensure that the sample arrives at the laboratory at an acceptable temperature.

10. Review the information on the pre-printed carrier shipping airbill (e.g., FedEx airbill) provided with the sampling supplies, and ensure that the sample is delivered to the FSIS Eastern Laboratory. Enter the return address information on the airbill.

11. Place the completed sample form and any unused sample seals in the container.

12. Insert the foam plug and press down to minimize the space between the sample and foam plug. If the shipping container does not have a foam plug, place the insulated lid on the container. Do not overfill the shipping container.

**NOTE:** Do not tape or wrap the samples or use any newspaper or similar material as packing material. Use of such materials may result in the sample being discarded by the laboratory.

13. Complete the information on the large bar-coded seal from the same FSIS Form 7355-2A/2B sample seal set, sign the seal, and affix the signed, large bar code seal across the seam of the closed sample box flap.

   a. For shipping containers with self-sticking closures, apply the seal across the closed inner flap of the box parallel to the edge of the closed flap. Then close the outer flap over the seal.

   b. For shipping containers without self-sticking closures, apply the seal across the closed outer flaps. Fasten the outer flaps with clear packaging tape.

14. Affix the carrier shipping airbill on the shipping container and remove any old stamp receipts and carrier shipping bar codes from the container.

15. Ensure that the sample remains under FSIS control until pickup by the FedEx.

D. The PHV is to ensure that the sample container lid is securely closed prior to shipping. If the sample container is leaking upon arrival, the laboratory may discard the sample. The PHV is to avoid storing shipping containers near heaters or in areas exposed to excessive heat.
E. Returning unused sample supplies

1. The PHV is to hold any unused sample supplies for future NARMS sampling tasks unless otherwise advised that no additional NARMS sampling tasks will be assigned to the establishment.

2. If at any time there is a need to return unused sampling supplies, the PHV is to send a request for a FedEx Ground shipping label by email through the FSIS - Sampling Supplies – Eastern Lab mailbox in Outlook.

3. The PHV is to include the establishment name, establishment number, mailing address, and project code in the email request. The PHV is to return any unused sampling supplies for the FSIS NARMS sampling program via ground shipping to the following address:

   USDA/FSIS/Eastern Laboratory
   Attn: NARMS-Microbiology
   Russell Research Center
   950 College Station Rd.
   Athens, GA 30605-2720

XI. SAMPLE RESULTS

Results of individual samples collected will not be posted in LIMS-Direct or PHIS. Individual establishment results will not be published. Sample results from the FSIS NARMS sampling program are non-regulatory; therefore, establishments do not need to hold products.

XII. QUESTIONS

Refer questions regarding this directive to the Risk, Innovations, and Management Staff through askFSIS, or by telephone at 1-800-233-3935 (press option 4). When submitting a question, use the Submit a Question tab, and enter the following information in the fields provided:

- **Subject Field:** Enter **Directive 10,100.1**
- **Question Field:** Enter your question with as much detail as possible.
- **Product Field:** Select **General Inspection Policy** from the drop-down menu.
- **Category Field:** Select **Sampling-General** from the drop down menu.
- **Policy Arena:** Select **Domestic (U.S.) Only** from the drop-down menu.

When all fields are complete, press **Continue** and at the next screen press **Finish Submitting Question**.

[Signature]

Assistant Administrator
Office of Policy and Program Development