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Title: FSIS Laboratory Regulatory Sample Pathogen Methods Table and Definitions			
Revision: .06	Replaces: .05	Effective: 03/07/22	

# **Laboratory Guidebook Notice of Change**

Chapter new, <u>revised</u>, or archived: MLG Appendix 3.06

Title: FSIS Laboratory Regulatory Sample Pathogen Methods Table and Definitions

Effective Date: 03/07/22

Description and purpose of change(s):

This Appendix was revised to reflect an update of the reporting time for *Campylobacter jejuni/coli/lari* negative and positive results. The reporting time was reduced from 3 days to 2 days for a screened negative result, and from 5 days to 4 days for a confirmed positive result.

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#### APP 3.1 Introduction

This appendix summarizes key steps of the Microbiology Laboratory Guidebook (MLG) 4 Salmonella, MLG 8 Listeria monocytogenes, MLG 5C Shiga toxin-producing Escherichia coli (STEC), and MLG 41 Campylobacter jejuni/coli/lari methods. The terminology used for describing method results are defined in the glossary.

Table 1 lists the screening technology and result reporting timeline for each method. On Day 1, regulatory samples arrive in the laboratory. Days listed for each analyte does not include delays (e.g., restreak for purity, waiting for completion of all analyses, rare strains requiring additional testing). Turnaround for these delays could include up to an additional 13 days. All confirmed positive isolates are forwarded to the Microbiology Characterization Branch (MCB) for further testing and upon completion are maintained in long-term storage. Table 1 does not include further characterization testing, e.g., Whole Genome Sequencing (WGS) and/or antimicrobial resistance (AMR) analyses.

AMR analyses are also performed on *Salmonella*, STEC, and *Campylobacter* as part of the National Antimicrobial Resistance Monitoring System (NARMS). This testing provides continuous monitoring of AMR profiles and allows detection of novel and emerging phenotypes and/or genotypes.

FSIS Laboratories utilize WGS data to infer the Salmonella serotype.

WGS is performed on all isolates and has replaced PFGE as the primary tool for determining source attribution and relatedness to other clinical, food, and environmental isolates. Sequence data is uploaded to the National Center for Biotechnology Information's (NCBI) website and is available to the public. Tools on the site compare the sequences to all uploads and ascertain relatedness and identify genes of interest. The data is also uploaded to CDC's PulseNet for further sharing and analysis with Federal and State public health partners.

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Table 1. Summary of Laboratory Regulatory Sample Pathogen Methods for STEC, L. monocytogenes, Salmonella, and Campylobacter.

Analyte	Microbiology	Screen Test	Confirmatory Tests	Days to	Days to	Days to	Days to
	Laboratory		(following culturing on tube and plating media;	Reporting:	Reporting:	Reporting:	Reporting: Final
	Guidebook		and following Immuno-Magnetic Separation	Screen	Potential +	Presumptive	+ Result
	Chapter #		[IMS] bead capture for STEC)	Negative	Result	+ Result	
STEC	MLG 5C	iQ-Check VirX and	O group serological and genetic confirmation:	Day 2	Day 2	Day 3	Day 4
		iQ-Check SerO II	Latex agglutination test kit and Real-Time PCR		•	•	·
Based on		Detection Assay			(Limited		
specifications of		(stx, eae, O group)	Isolate confirmation:		distribution)		
MT sampling			Bruker® MALDI Biotyper		distribution)		
project			Real-Time PCR Shiga toxin genes confirmation:				
			stx and eae genes; if needed for inconclusive results, WGS				
Listeria	MLG 8	3M Molecular	Observation of β-hemolytic colonies	D 2	NT A	D 4 5	D 5 (
	WILG 6	Detection Assay	Observation of p-hemorytic colonies	Day 3	NA	Day 4-5	<b>Day 5-6</b>
monocytogenes		Detection Assay	Isolate confirmation:				
			Bruker® MALDI Biotyper;				
			Genetic Identification Testing, if needed, for				
			speciation – WGS				
Salmonella spp.	MLG 4	3M Molecular	speciation was	Day 2	NA	Day 5	Day 5-6
Samonena spp.	1.120	Detection Assay	Isolate confirmation:	Day 2	11/1	NA for	Day 3-0
		Detection rissay	Bruker® MALDI Biotyper				
C 1.1 .	MI C 41	2) ( ) ( ) 1		<b>D</b> 0	<b>N.</b> T. A.	HACCP	<b>D</b> 4
Campylobacter	MLG 41	3M Molecular	Typical colonies subject to same day confirmation:	Day 2	NA	NA	Day 4
jejuni/coli/lari		Detection Assay	Bruker® MALDI Biotyper				
for Enrichment							
method							

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### **APP 3.2 Definitions:**

Potential positive STEC – Enrichment medium from a sample which yields a positive when screened on real-time PCR for each of the targeted genes (*eae*, *stx*1/2, and O group) of one or more of seven serogroups (O26, O45, O103, O111, O121, O145, O157).

Presumptive positive STEC – One or more typical colonies on modified Rainbow agar (mRBA) which agglutinate with latex agglutination reagents and are positive on real-time PCR for the *eae*, *stx*, and O group of one or more of the seven serogroups.

Confirmed positive STEC – One or more isolates from Sheep Blood Agar (SBA) positive on real-time PCR for the *eae*, *stx*, and O group of one or more of seven serogroups, and identified as *Escherichia coli*.

Presumptive positive *L. monocytogenes* – A sample from which one or more typical colonies produces beta-hemolysis on Horse Blood Overlay Agar (HBO).

Confirmed positive *L. monocytogenes* – A beta-hemolytic isolate is identified as *L. monocytogenes*.

Presumptive positive Salmonella spp. – A sample yields one or more isolates which show typical appearance on Triple Sugar Iron (TSI) and Lysine Iron Agar (LIA) slants.

Confirmed positive *Salmonella* spp. – *Salmonella* typical colonies is identified as *Salmonella* spp. All *Salmonella* isolates are forwarded for further characterization, e.g., serotype determination, WGS and AMR analyses.

Confirmed positive *Campylobacter* spp. – *Campylobacter* typical colonies is identified as *C. jejuni, C. coli, and/or C. lari.* All *Campylobacter* isolates are forwarded for further characterization, e.g., WGS and AMR analyses.